



http://intl.elsevierhealth.com/journals/mehy

## Any beneficial effects of mycobacteria on multiple sclerosis and experimental autoimmune encephalitis may include stimulation of the sympathetic nervous system

### Shahram Shahabi <sup>a</sup>,\*, Zuhair Muhammad Hassan <sup>b</sup>, Nima Hosseini Jazani <sup>a</sup>

<sup>a</sup> Department of Microbiology and Immunology, Medical Faculty, Urmia Medical Sciences University, Road of Nazloo, Urmia, Iran <sup>b</sup> Department of Immunology, Faculty of Medical Sciences, Tarbiat Modarres University, Tehran, Iran

Received 12 November 2005; accepted 5 January 2006

Summary The inhibitory effects of mycobacterial infection and mycobacterium components on multiple sclerosis (MS) and experimental autoimmune encephalitis (EAE; an animal model for MS) have been known for years. However, this effect seems like a paradox that both mycobacterial infection and MS induce type I immune responses. Some mechanisms have been proposed or even proven for this effect in different studies, but among them there is no hint of a possible role for the nervous system (NS). Regarding the close relations between sympathetic nervous system (SNS) and MS disease course, it can be hypothesized that SNS may have a role in the effects of mycobacterium on MS. Hypothesis: SNS can be stimulated by pro-inflammatory cytokines such as TNF- $\alpha$  and IL1- $\beta$ , production of which are induced by mycobacterial infection or mycobacterium components. Although these cytokines can inhibit SNS in the site of inflammation caused by mycobacterium, they increase sympathetic tone in other places. The beneficial role of SNS in inhibiting or attenuating the course of MS and EAE has been suggested. Inhibitory effects of stimulated SNS on MS may occur via different ways such as inhibiting the production of pro-inflammatory cytokines and inducing the synthesis of anti-inflammatory cytokines, in other words, shifting the immune responses from type 1 toward type 2, as well as, induction of suppressor/regulator T lymphocytes, induction of heat shock proteins in brain and increasing the expression of Fas and Fas-ligand. Therefore, it seems that stimulation of SNS by mycobacterial infection or mycobacterium components is a key step in the mechanism of beneficial effects of mycobacterium on MS. © 2006 Elsevier Ltd. All rights reserved.

\* Corresponding author. Address: P.O. Box 14115-183, Tehran, Iran. Tel.: +98 2188011001x3565; fax: +98 2188013030.

*E-mail addresses*: shahabsh@modares.ac.ir, shahabirabori@ yahoo.com (S. Shahabi).

### Introduction

It has been demonstrated that infectious agents contribute to the development of autoimmune diseases [1] including multiple sclerosis (MS) [2]. However, there is evidence indicating that infection

0306-9877/\$ - see front matter  $\, \textcircled{0}$  2006 Elsevier Ltd. All rights reserved. doi:10.1016/j.mehy.2006.01.021

diseases could have a protective effect on autoimmunity. In particular, it has been shown that mycobacterial infection and mycobacterium components can modulate MS and experimental autoimmune encephalomyelitis (EAE; an animal model for multiple sclerosis) in mice [3-8] through some mechanisms such as deletion of auto reactive T cells, induction of suppressor (regulator) T cells and redirected trafficking of activated auto antigen-specific T cells to the local inflammatory sites, induced by mycobacterial infection [3,6-11].

To support the beneficial effects of Bacillus Calmet-Guerin (BCG) on human autoimmunity, an inverse relationship between incidence of MS and spontaneous positive tuberculin skin tests was reported by Andersen et al. [12]. They suggested that early mycobacterial infection may be protective against MS. BCG vaccine has been used in recent clinical trials against MS and resulted in a 51% reduction in lesions, demonstrated by gadolinium-enhanced magnetic resonance imaging [13].

The beneficial effects of mycobacterium on MS are an apparent paradox that a Th1-promoting immune stimulus can have a useful effect on a purported Th1-mediated autoimmune disease [7].

As mentioned, some mechanisms have been proposed or proven for the effects of mycobacterium on MS, but there is no hint to the possible role of nervous system. Regarding the close relations between sympathetic nervous system and MS course [14], it can be hypothesized that SNS may have a role in the effects of mycobacterium on MS.

Therefore, in the present paper we will discuss this hypothesis that sympathetic nervous system (SNS) may have a crucial role in the beneficial effects of mycobacterium and its components on MS. Mycobacterial infection or mycobacterium components can increase systemic sympathetic tone including sympathetic tone of central nervous system, leading to the attenuation of the MS disease course via the ways that will be mentioned in coming paragraphs.

### Mycobacterium components can stimulate the sympathetic nervous system

Mycobacterium components can induce local immune system cells to produce pro-inflammatory cytokines including TNF- $\alpha$ , IFN- $\gamma$ , IL-12, IL-1 $\beta$  and IL-6 [15–20]. These cytokines can inhibit local sympathetic tone, whereas they increase systemic sympathetic tone [21]. Therefore, what appears during the infection by mycobacterium is the reduction of sympathetic tone in the infection loci and its increase in other places including CNS.

Any immune challenge that threatens the stability of the internal milieu can be regarded as a stressor, i.e. under certain conditions; an immune response can activate the stress system. It seems that an inflammatory/immune response to mycobacterium components may actually activate SNS, as other stressors or stimuli do [21]. It has been shown that IFN- $\alpha$ , which can be induced by mycobacterium components [17], TNF- $\alpha$ , IL-1 (especially IL-1 $\beta$ ) and IL-6 can signal the brain to trigger the activation of both SNS and Hypothalamus-Pituitary-Adrenal (HPA) axis through a complex Corticotropin-releasing Hormone (CRH)-dependent pathway [21]. Thus, the SNS, similar to HPA axis [21], is involved also in a long feedback loop between lymphoid organs and CNS. The afferent limb of this loop seems to be operated by blood-borne cytokines, which activate the central components of the stress system via circulation or through the vagus nerve afferents. The efferent loop consists of the SNS, its projections to different organs and the release of NE from the sympathetic nerve terminals in these organs [21].

Although the above-mentioned cytokines trigger centrally the sympathetic output, which results in an increase of NE turn over in several organs, it has been shown that TNF- $\alpha$  and IL-1 inhibit SNS activation in the place of administration [21], so the local effect of these cytokines might be absolutely different. Therefore, it seems that mycobacterium infection and mycobacterial components can decrease sympathetic tone via inducing the above-mentioned cytokines production in the inflammation loci.

## Effect of increased sympathetic tone on the pathogenesis of MS

# Suppression of inflammatory molecules production and shifting the immune system from type 1 toward type 2

There are many indications showing that NE and epinephrine, inhibit the production of type 1/proinflammatory cytokines, such as interleukin 12 (IL-12), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interferon- $\gamma$  (IFN- $\gamma$ ) by antigen-presenting cells and T helper1 (Th1) cells through stimulation of the  $\beta_2$ adrenoreceptor-cAMP-protein kinase A pathway, whereas they stimulate the production of antiinflammatory/Th2 and Th3 cytokines such as interleukin 10 (IL-10) and transforming growth factor- $\beta$  (TGF- $\beta$ ). Through this mechanism, endogenous catecholamines may act systemically to cause a selective suppression of Th1 responses and cellular immunity, and result in a Th2 shift toward the dominance of humoral immunity [21]. Stimulation of SNS seems to pose similar effects on CNS.

Now, it has been clearly demonstrated that Nor-Adrenalin (NA) can suppress the expression of inflammatory molecules, including adhesion molecules, class II MHC molecules and NOS, in both glial and endothelial cells [22]. It has also been shown that the capacity of both astroglial and microglial cells to synthesize the pro-inflammatory (IL-1 $\beta$ and TNF- $\alpha$ ) cytokines, is potently reduced by NA. There are many other studies suggesting that activation of  $\beta$ -adrenergic receptors on astrocytes can inhibit inflammatory molecules production and ameliorate EAE [14].

Furthermore, it is known that there is a shift toward type 1 immune responses in MS disease and that the shifting of the immune responses toward type 2 can attenuate the course of the disease [5]. For instance, it has been shown that the protective effect of Schistosomiasis on EAE and MS is due to the shifting of the immune responses toward type 2 in Schistosomiasis [23].

### Induction of heat shock proteins

All cells, from bacteria to human, show a common intricate response to stress, which protects them from injury. Heat shock proteins (HSPs), also known as stress proteins and molecular chaperones, play a central role in this response and procellular homeostatic processes tect from environmental and physiological insults [24]. Heat shock proteins (HSPs) act as molecular chaperones and/or have anti-apoptotic activities under physiological conditions. Expression of two heat shock proteins namely HSP70 and HSP27 in the brain is notable as they are highly inducible in glial cells and neurons following a wide range of stressors including ischemia, epileptic seizure and hyperthermia. Both of these HSPs have neuroprotective effects on the brain and spinal cord due to their anti-apoptotic and chaperoning activities [25]. Accumulation of these HSPs, whether induced physiologically, pharmacologically or genetically, or by direct administration of the proteins, is known to protect the brain and spinal cord from neurodegenerative diseases [24-26].

Stimulation of SNS can act as a sublethal stress and induce HSPs in various organs including the brain. Stimulation of adrenoreceptors can induce HSP-70 expression in the brain cells [27–30] and protect these cells from neurodegenerative diseases, including MS. While the mechanism of this protection has largely been thought to be due to chaperoning and anti-apoptotic functions of HSPs, it has been also shown that these proteins may directly interfere with inflammation [31]. According to these findings, some anti-inflammatory effects of SNS on the brain may be explainable via induction of HSPs.

### Other effects of increased sympathetic tone on immunopathogenesis of MS disease

It has been shown that stimulation of SNS can induce regulatory (suppressor) T lymphocytes and attenuate immune responses. Different adrenoreceptors are known as molecules responsible for the induction of regulatory T lymphocytes and increasing the T regulatory (suppressor) vs. T helper and T regulatory (suppressor) vs. T cytolytic ratios [32,33].

The role of regulatory T lymphocytes in the effects of mycobacterium tuberculosis components on EAE has been demonstrated [7,9].

It has been shown that the number of Gamma Delta T lymphocytes and their activity increases due to the activation of SNS [34,35]. There are some evidences that Gamma Delta lymphocytes play an important role in attenuating the course of EAE [36]. In addition, production of TGF- $\beta$  by Gamma Delta T cells has been suggested as a potential mechanism in mycobacterium-induced protection against EAE [7].

Another effect of increased sympathetic tone is the increase of activated T lymphocytes apoptosis. It has been shown that catecholamines can inhibit T- and B-lymphocytes activities, mediated via induction of Bcl-2/Bax and Fas/FasL involved apoptosis [37]. This finding indicates a mechanism for the regulation of lymphocytes activity in the central nervous system, whereby elevated regional levels of catecholamines might lead to the apoptosis of auto-reactive lymphocytes [36] and immunoprivilege of the brain [37].

It has been also shown that catecholamines can inhibit the function of effector cytotoxic T lymphocytes [21] involved in the pathogenesis of MS [38].

The activity of auto-reactive T lymphocytes can be directly affected by increased sympathetic tone. Bedoui et al. [39] have shown that neuropeptide Y(NPY) significantly inhibits myelin oligodendrocyte glycoprotein (MOG) specific Th1 lymphocytes in EAE. They showed that these auto-reactive T cells, which have a key role in pathogenesis of EAE, are a major target for NPY [39].

### Discussion

The present paper suggested stimulation of SNS and sympathetic tone increase may be one of the possible mechanisms for the inhibition or attenuation of EAE and MS disease courses, caused by mycobacterial infection and mycobacterium components.

The beneficial role of SNS in inhibiting or attenuating the course of MS and EAE has been suggested. Chemical sympathectomy with 6-hydroxydopamine (6-OHDA) at birth, led to a more severe course of the disease in an EAE rat model, as well as in an adoptive transferred EAE [40,41]. In a study with secondary progressive course MS (SPMS) patients, Makhlouf et. al. [42] showed that after 14 days of treatment with Salbutamol, the in vitro production of IL-12 by monocytes and dendritic cells significantly decreases compared to baseline, with persisting effects for, at least, 1 week after the treatment. Furthermore, they reported an increase of Th2 cytokines [43]. Based on these findings, a randomized trial of add-on treatment of Salbutamol to glatiramer acetate is under progress [14].

As mentioned above, it is absolutely possible that mycobacterium components can stimulate SNS through production of specific pro-inflammatory cytokines. The finding that IFN- $\gamma$  or TNF- $\alpha$  has protective effects on EAE models, when administered late during the disease process [3], supports this hypothesis that induction of pro-inflammatory cytokines production by mycobacterium may has a key role in the protective effects of mycobacterium on EAE and MS.

The suggested mechanisms of the beneficial effects of mycobacterium components on EAE models and MS disease can also be related to the stimulation of SNS. These mechanisms include the induction of suppressor T lymphocytes and inhibition of auto-reactive T lymphocytes, induced by mycobacterium [3,6–9]. As mentioned above, both induction of suppressor T lymphocytes and inhibition of auto-reactive T lymphocytes can be caused by increase in sympathetic tone [32,33,39]. It has also been suggested that redirected trafficking of activated auto antigen-specific T cells to local inflammatory sites, induced by mycobacterial infection, is one of the mechanisms, through which BCG infection can ameliorate the EAE course [7]. As SNS can influence lymphocyte trafficking and recirculation [21], it may has an effect on the trafficking of activated auto antigen-specific T cells. As mentioned above, the sympathetic tone may decrease in local inflammatory sites, induced by mycobacterial infection, and increase in other places including CNS. Therefore, stimulation of SNS can be one of the first steps in the mechanism of Mycobacterium effects on MS disease course, and other proven or suggested mechanisms can be due to this increased sympathetic tone.

In summary, it has been hypothesized that stimulation of SNS by mycobacterial infection or Mycobacterium components and other possible effects of increased sympathetic tone on CNS immune and non-immune cells may have an important role in beneficial effects of mycobacterium on MS disease course, whereas interaction of other mechanisms is highly possible.

This hypothesis could be tested via different ways for example according this hypothesis a chemical sympathectomy should inhibit the beneficiary effects of BCG on EAE.

#### References

- Wucherpfennig KW. Mechanisms for the induction of autoimmunity by infectious agents. J Clin Invest 2001;108: 1097–104.
- [2] Hunter SF, Hafler DA. Ubiquitous pathogens: links between infection and autoimmunity in MS? Neurology 2000;55: 164–5.
- [3] Christen U, von Herrath MG. Infections and autoimmunity good or bad? J Immunol 2005;174:7481–6.
- [4] Kamradt T, Goggel R, Erb KJ. Induction, exacerbation and inhibition of allergic and autoimmune diseases by infection. Trends Immunol 2005;26:260–7.
- [5] Sewell DL, Reinke EK, Hogan LH, et al. Immunoregulation of CNS autoimmunity by helminth and mycobacterial infections. Immunol Lett 2002;82:101–10.
- [6] Ben-Nun A, Mendel I, Sappler G, et al. A 12 kDa protein of mycobacterium tuberculosis protects mice against experimental autoimmune encephalomyelitis. Protection in the absence of shared T cell epitopes with encephalitogenic proteins. J Immunol 1995;154:2939–48.
- [7] Sewell DL, Reinke EK, Co DO, et al. Infection with mycobacterium bovis BCG diverts traffic of myelin oligodendroglial glycoprotein autoantigen-specific T cells away from the central nervous system and ameliorates experimental autoimmune encephalomyelitis. Clin Diagn Lab Immunol 2003;10:564–72.
- [8] Ben-Nun A, Yossefi S, Lehmann D. Protection against autoimmune disease by bacterial agents. II. PPD and pertussis toxin as proteins active in protecting mice against experimental autoimmune encephalomyelitis. Eur J Immunol 1993;23:689–96.
- [9] Grabie N, Wohl I, Youssef S, et al. Expansion of neonatal tolerance to self in adult life. I. The role of a bacterial adjuvant in tolerance spread. Int Immunol 1999;11(6):899–906.
- [10] Constant P, Davodeau F, Peyrat MA, et al. Stimulation of human gamma delta T cells by non-peptidic mycobacterial ligands. Science 1994;264:267–70.
- [11] Muller D, Pakpreo P, Filla J, et al. Increased gamma-delta T-lymphocyte response to mycobacterium bovis BCG in major histocompatibility complex class I-deficient mice. Infect Immun 1995;63:2361–6.
- [12] Andersen E, Isager H, Hyllested K. Risk factors in multiple sclerosis: tuberculin reactivity, age at measles infection,

tonsillectomy and appendectomy. Acta Neurol Scand 1981;63:131-5.

- [13] Ristori G, Buzzi MG, Sabatini U, et al. Use of Bacille Calmette-Guerin (BCG) in multiple sclerosis. Neurology 1999;53:1588–9.
- [14] Gold SM, Mohr DC, Huitinga I, et al. The role of stressresponse systems for the pathogenesis and progression of MS. Trends Immunol 2005(October 6) [Epub ahead of print].
- [15] Valone SE, Rich EA, Wallis RS, et al. Expression of tumor necrosis factor in vitro by human mononuclear phagocytes stimulated with whole Mycobacterium bovis BCG and mycobacterial antigens. Infect Immun 1988;56:3313–5.
- [16] Peetermans WE, Raats CJ, Langermans JA, et al. Mycobacterial heat-shock protein 65 induces proinflammatory cytokines but does not activate human mononuclear phagocytes. Scand J Immunol 1994;39:613–7.
- [17] Koh YI, Choi IS, Lee JJ. Effects of cytokine milieu secreted by BCG-treated dendritic cells on allergen-specific Th immune response. J Korean Med Sci 2004;19:640–6.
- [18] Zhang Y, Doerfler M, Lee TC, et al. Mechanisms of stimulation of interleukin-1 $\beta$  and tumor necrosis factor- $\alpha$  by mycobacterium tuberculosis components. J Clin Invest 1993;91:2076–83.
- [19] Barnes PF, Chatterjee D, Abrams JS, et al. Cytokine production induced by mycobacterium tuberculosis lipoarabinomannan. Relationship to chemical structure. J Immunol 1992;149:541–7.
- [20] Peetermans WE, Raats CJ, van Furth R, et al. Mycobacterial 65-kilodalton heat shock protein induces tumor necrosis factor  $\alpha$  and interleukin 6, reactive nitrogen intermediates, and toxoplasmastatic activity in murine peritoneal macrophages. Infect Immun 1995;63:3454–8.
- [21] Elenkov IJ, Wilder RL, Chrousos GP, et al. The sympathetic nerve – an integrative interface between two supersystems: the brain and the immune system. Pharmacol Rev 2000;52:595–638.
- [22] Feinstein DL, Heneka MT, Gavrilyuk V, et al. Noradrenergic regulation of inflammatory gene expression in brain. Neurochem Int 2002;41:357–65.
- [23] La Flamme AC, Ruddenklau K, Backstrom BT. Schistosomiasis decreases central nervous system inflammation and alters the progression of experimental autoimmune encephalomyelitis. Infect Immun 2003;71:4996–5004.
- [24] Tytell M, Hooper PL. Heat shock proteins: new keys to the development of cytoprotective therapies. Exp Opin Ther Targets 2001;5:267–87.
- [25] Franklin TB, Krueger-Naug AM, Clarke DB, et al. The role of heat shock proteins Hsp70 and Hsp27 in cellular protection of the central nervous system. Int J Hyperthermia 2005;21:379–92.
- [26] Johnson JD, Campisi J, Sharkey CM, et al. Adrenergic receptors mediate stress-induced elevations in extracellular Hsp72. J Appl Physiol 2005;99:1789–95.
- [27] Murphy SJ, Song D, Welsh FA, et al. The effect of hypoxia and catecholamines on regional expression of heat-shock protein-72 mRNA in neonatal piglet brain. Brain Res 1996;727:145–52.
- [28] Udelsman R, Li DG, Stagg CA, et al. Adrenergic regulation of adrenal and aortic heat shock protein. Surgery 1994;116(2):177-82.

- [29] Johnson JD, Campisi J, Sharkey CM, et al. Adrenergic receptors mediate stress-induced elevations in extracellular Hsp72. J Appl Physiol 2005;99(5):1789–95 [Epub 2005 Jul 21].
- [30] Meng X, Brown JM, Ao L, et al. Norepinephrine induces cardiac heat shock protein 70 and delayed cardioprotection in the rat through  $\alpha$  1 adrenoceptors. Cardiovasc Res 1996;32:374–83.
- [31] Heneka MT, Gavrilyuk V, Landreth GE, et al. Noradrenergic depletion increases inflammatory responses in brain: effects on IκB and HSP70 expression. J Neurochem 2003;85:387–98.
- [32] Murray DR, Irwin M, Rearden CA, et al. Sympathetic and immune interactions during dynamic exercise. Mediation via a  $\beta$  2-adrenergic-dependent mechanism. Circulation 1992;86:203–13.
- [33] Cao L, Hudson CA, Lawrence DA. Immune changes during acute cold/restraint stress-induced inhibition of host resistance to Listeria. Toxicol Sci 2003;74: 325–34.
- [34] Suzuki S, Toyabe S, Moroda T, et al. Circadian rhythm of leucocytes and lymphocytes subsets and its possible correlation with the function of the autonomic nervous system. Clin Exp Immunol 1997;110:500–8.
- [35] Minagawa M, Narita J, Tada T, et al. Mechanisms underlying immunologic states during pregnancy: possible association of the sympathetic nervous system. Cell Immunol 1999;196:1–13.
- [36] Ponomarev ED, Dittel BN. Gamma delta T cells regulate the extent and duration of inflammation in the central nervous system by a Fas ligand-dependent mechanism. J Immunol 2005;174:4678–87.
- [37] Bergquist J, Josefsson E, Tarkowski A, et al. Measurements of catecholamine-mediated apoptosis of immunocompetent cells by capillary electrophoresis. Electrophoresis 1997;18(10):1760-6.
- [38] Tsuchida T, Parker KC, Turner RV, et al. Autoreactive CD8+ T-cell responses to human myelin protein-derived peptides. Proc Natl Acad Sci USA 1994;91(23):10859–63 [Erratum in: Proc Natl Acad Sci USA 1995;92:9432].
- [39] Bedoui S, Miyake S, Lin Y, et al. Neuropeptide Y (NPY) suppresses experimental autoimmune encephalomyelitis: NPY1 receptor-specific inhibition of autoreactive Th1 responses in vivo. J Immunol 2003;171:3451–8.
- [40] Chelmicka-Schorr E, Checinski M, Arnason BG. Chemical sympathectomy augments the severity of experimental allergic encephalomyelitis. J Neuroimmunol 1988;17: 347–50.
- [41] Chelmicka-Schorr E, Kwasniewski MN, Wollmann RL. Sympathectomy augments adoptively transferred experimental allergic encephalomyelitis. J Neuroimmunol 1992;37: 99–103.
- [42] Makhlouf K, Comabella M, Imitola J, et al. Oral salbutamol decreases IL-12 in patients with secondary progressive multiple sclerosis. J Neuroimmunol 2001;117:156– 65.
- [43] Makhlouf K, Weiner HL, Khoury SJ. Potential of β2-adrenoceptor agonists as add-on therapy for multiple sclerosis: focus on salbutamol (albuterol). CNS Drugs 2002;16:1–8.

Available online at www.sciencedirect.com