3 4

5 6

7 8

9

10

11

12 13

14

16

17

DOI: 10.1111/myc.12716

<u>ORIGINALARTICLE</u>

WILEY

In vitro antifungal activity of amphotericin B and 11 comparators against *Aspergillus terreus* species complex

¹Department of Medical Mycology and Parasitology, Invasive Fungi Research Center, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran

Afsane Vaezi^{1,2} | Hamed Fakhim^{3,4} | Amir Arastehfar⁵ | Tahereh Shokohi¹ .

Mohammad T. Hedayati¹ | Sadegh Khodavaisy⁶ | Ali Rezaei-Matehkolaei⁷ | Parisa Badiee⁸ | Ferry Hagen⁹ | Cornelia Lass-Flörl¹⁰ | Eric Dannaoui¹¹ | Jacques F. Meis ^{9,12} | Hamid Badali ¹⁰

²Student Research Committee Center, Mazandaran University of Medical Sciences, Sari, Iran ³Department of Medical Parasitology and Mycology, Faculty of Medicine, Urmia University of Medical Sciences, Urmia, Iran

20 ⁴Cellular and Molecular Research Center, Urmia University of Medical Sciences, Urmia, Iran

Revised: 23 September 2017

⁵Westerdijk Fungal Biodiversity Institute, Utrecht, The Netherlands

- 22 ⁶Department of Medical Mycology and Parasitology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran
- 23 ⁷Infectious and Tropical Diseases Research Center, Health Research Institute, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran
- 24 ⁸Alborzi Clinical Microbiology Research Center, Shiraz University of Medical Sciences, Shiraz, Iran
- 25 ⁹Department of Medical Microbiology and Infectious Diseases, Canisius-Wilhelmina Hospital(CWZ), Nijmegen, The Netherlands

Summary

- 26 ¹⁰Division of Hygiene and Medical Microbiology, Medical University of Innsbruck, Innsbruck, Austria
- 27 ¹¹Faculté de Médecine, APHP, Université Paris-Descartes, Hôpital Européen Georges Pompidou, Unité de Parasitologie-Mycologie, Paris, France
- 28 ¹²Center of Expertise in Mycology Radboudumc, CWZ, Nijmegen, The Netherlands

29 30

- 31 Correspondence
- 32 Hamid Badali, Department of Medical
- Mycology and Parasitology, Invasive Fungi
- 33 Research Center, School of Medicine,
- 34 Mazandaran University of Medical Sciences,
- 35 Sari, Iran.
- Email: badalii@yahoo.com

37 Funding information

- 38 Mazandaran University of Medical Sciences, Grant/Award Number: 2206
- 39 40
- 41
- 42 43
- 45 46 47

48 49

50

51 52 53

44

of amphotericin B and 11 comparators against clinical (n = 36) and environmental (n = 45) A. *terreus* isolates. In vitro antifungal susceptibility was performed using the

Aspergillus terreus infections are difficult to treat because of the intrinsic resistance to

amphotericin B, and higher mortality compared to infections caused by other Aspergillus

species. The aim of the present study was to determine the in vitro antifungal activity

CLSI M38--A2 procedure. Amphotericin B exhibited the highest MICs (MIC range, 0.125--4 μ g/mL; MIC90, 2 μ g/mL), followed by terbinafine (MIC range, 0.002--1 μ g/mL; MIC90, 1 μ g/mL). Only one isolate (1/81) showed amphotericin B MIC above the epidemiologic cut--off value (ECV; 4 μ g/mL). None of the isolates had a MIC of>ECV for voriconazole, itraconazole and posaconazole. The reasons for the difference in amphotericin B susceptibility patterns between studies remain unknown. The genetic and species diversity, clinical, environmental and ecological factors in *Terrei* section on various amphotericin B susceptibility profiles in different countries should be considered more as the main reasons associated with these differences.

KEYWORDS

amphotericin B, Aspergillus terreus species complex, In vitro antifungal activity, Iran

2

3 The emergence of drug resistance in fungal infections is one of the most significant epidemiological changes in current decades.¹ Thus. 4 therapeutic options available for the treatment of these infections 5 have become limited.² Amphotericin B is generally considered the 6 7 mainstay of treatment of severe fungal infections. Invasive fungal 8 infections due to Aspergillus terreus are difficult to treat because 9 of the intrinsic resistance to amphotericin B, and higher mortality compared to infections caused by other Aspergillus species.³ The in 10 vitro and in vivo amphotericin B resistance in A. terreus isolates is 11 12 not very well known so far. However, it may be correlated to less absorption and amphotericin B to cause cell death by oxidative dam-13 age.⁴ In addition, upregulation of ERG5, ERG6 and ERG25 genes 14 15 (ergosterol biosynthesis genes) could be responsible for decreased 16 ergosterol content in the cell membrane leading to the development of resistance.^{4,5} Clinical studies have shown a lack of response 17 18 to amphotericin B and variable efficacy of azole therapy, particu-19 larly for voriconazole, with a high percentage of treatment failure in *A. terreus* infections.^{6,7} Moreover, alterations in *cyp51A* such as 20 21 the M217I mutation are responsible for acquired voriconazole resistance in A. terreus.⁸ In many countries of the world, there are no 22 23 epidemiological data about the antifungal susceptibility profiles of 24 A. terreus available. The aim of the present study was to determine 25 the in vitro antifundal activity of amphotericin B and 11 compara-26 tors against 81 clinical and environmental A. terreus isolates from 27 different geographical regions in Iran. 28

29

30 2 | MATERIALS AND METHODS

31

32 In all, 81 clinical and environmental of *A. terreus* isolates were ob-33 tained from Sari (36.5659°N, 53.0586°E), Tehran (35.6892°N,

34 51.3890°E), Ahvaz (31.3183°N, 48.6706°E), Mashhad (36.2605°N, 59.6168°E) and Shiraz (29.5918°N, 52.5837°E). The collection con-35 sisted of 36 clinical isolates from a variety of specimens comprising, 36 bronchoalveolar lavage fluid (n = 7), biopsy (n = 3), pharynx (n = 3), 37 38 sinus (n = 1), sputum (n = 8) and nail (n = 14) (Table 1). In addition, 39 45 environmental isolates collected from air (n = 7) and soil (n = 38)samples in hospital surroundings were included (Table 1). Soil isolates 40 were processed as previously described by Chowdhary et al.⁹ and all 41 42 isolates were deposited at -70°C in the reference culture collection of Invasive Fungi Research Center (IFRC), Mazandaran University of 43 44 Medical Sciences, Sari, Iran. All isolates were identified to species level 45 by DNA sequencing of partial β -tubulin gene, using primers Bt2a (5 `--GGTAACCAAATCGGTGCTGCTTTC--3`) and Bt2b (5`--ACCCTCAG 46 TGTAGTGACCCTTGGC--3`) as previously described by Hong et al¹⁰ 47 In vitro antifungal susceptibility was performed using the CLSI M38--48 A2 procedure.¹¹ Antifungal agents were dispensed into microdilution 49 trays at final concentration ranges of 0.016--16 µg/mL for ampho-50 51 tericin B (Bristol--Myers--Squib, Woerden, Netherlands), itraconazole 52 (Janssen Research Foundation, Beerse, Belgium), voriconazole (Pfizer, 53 Central Research, Sandwich, United Kingdom), posaconazole (Merck,

Whitehouse Station, NJ) and isavuconazole (Basilea Pharmaceuticals, Basel, Switzerland), 0.063--64 µg/mL for fluconazole (Pfizer, Groton, CT, USA), 0.001--1 µg/mL for lanoconazole and luliconazole (Nihon Nohvaku Co, Osaka, Japan), 0.004--4 µg/mL for terbinafine (Novartis Research Institute, Vienna, Austria), caspofungin (Merck Sharp & Dohme, Haarlem), micafungin (Astellas Pharma, Ibaraki, Japan) and anidulafungin (Pfizer, Central Research, Sandwich, United Kingdom). MICs were compared with previously published epidemiological cut-off values (ECVs) for AMB (4 µg/mL), ITC (1 µg/mL); VRC (1 µg/mL); POS (0.5 μg/mL); ISA (1 μg/mL) and CFG (0.25 μg/mL).^{12,13} Candida parapsilosis (ATCC 22019) and Candida krusei (ATCC 6258) reference strains were included as quality controls.¹¹ All tests were performed in duplicate and the Students t--test was used to compare MICs distribution between clinical and environmental A. terreus species complex isolates, with the SPSS statistical package (version 7.0), P values of <.05 were considered statistically significant.

2.1 | Patient details

The patients were primary included based on clinically suspected *Aspergillus* infection and confirmed by histopathology and mycology examination. Data on demographics, clinical characteristics, underly-ing condition, radiological features and mycological findings were col-lected with positive *A. terreus* cultures. The patients were categorised in four groups comprising, invasive aspergillosis (IA), aspergilloma, chronic pulmonary aspergillosis (CPA) and onychomycosis according to the diagnostic criteria.¹⁴⁻¹⁶ The patient without proven or prob-able disease was classified into the colonised group. This study was approved by the Ethics Commission of Mazandaran University of Medical Sciences, Sari, Iran (nr. 2206).

3 | RESULTS

Molecular identification showed that 66 and 15 isolates were A. ter-reus sensu stricto and A. citrinoterreus, respectively. 33.3% (5 of 15) and 26.7% (4 of 15) of A. citrinoterreus were isolated from sputum and soil, respectively. 69.7% of A. terreus sensu stricto were also isolated from soil (n = 33) and nail (n = 13) samples. However, other cryptic species of Aspergillus section Terrei were not detected. Nucleotide sequences of all isolates have been deposited in GenBank under the accession numbers MF185011 to MF185091. Table 2 summa-rises the MIC range, mode, geometric mean (GM) MIC, MIC50 and MIC90 for 12 antifungal drugs against 81 A. terreus isolates. Among the antifungal drugs, amphotericin B exhibited the highest MICs (MIC range, 0.125--4 µg/mL; MIC90, 2 μ g/mL), followed by terbinafine (MIC range, 0.002--1 μ g/mL; MIC90, 1 μg/mL), voriconazole (MIC range, 0.125--1 μg/mL; MIC90, 0.5 μg/mL), posaconazole (MIC range, 0.031--0.5 µg/mL; MIC90, 0.5 µg/mL) and itraconazole (MIC range, 0.016--2 µg/mL; MIC90, 0.25 µg/mL). However, the novel imidazoles, that is, lanoconazole (MIC range, 0.001--0.031; MIC90, 0.031 µg/mL) and luliconazole (MIC range, 0.001--0.031; MIC90, 0.008 µg/mL) dem-onstrated potent activity against all A. terreus isolates, in comparison

TABLE 1 Isolation data of Aspergillus terreus species complex in Iran

Species nr (%)	IFRC nr	Beta Tubulin accession nr	Source	Specimen	Age/Sex	Underlying condition	First-line therapy	Ori
A. terreus sensu	IFRC 1131	MF185016	Environmental	Soil				Teh
stricto, 66	IFRC 1132	MF185017	Clinical	Pharynx	15/Female	Cystic fibrosis	ІТС	Teh
(81.5%)	IFRC 1133	MF185018	Clinical	BAL	31/Female	Bilateral pulmonary echinococcosis	пс	Sar
	IFRC 1134	MF185019	Clinical	Nail	29/Male	Diabetes	ND	Shii
	IFRC 1135	MF185020	Clinical	Pharynx	21/Male	Cystic fibrosis	ITC	Teh
	IFRC 1136	MF185021	Clinical	Nail	62/Female	None	ND	Shii
	IFRC 1137	MF185022	Clinical	Pharynx	18/Male	Cystic fibrosis	ITC	Teh
	IFRC 1165	MF185023	Environmental	Soil		-		Sar
	IFRC 1166	MF185024	Environmental	Soil				Sar
	IFRC 1168	MF185025	Clinical	Biopsy (Lung)	52/Male	Tuberculosis pneumonia	ND	Teh
	IFRC 1169	MF185026	Clinical	BAL	48/Male	Kidney Transplantation	AMB/VRC	Teh
	IFRC 1171	MF185027	Clinical	Nail	25/Male	None	ND	Ahv
	IFRC 1172	MF185028	Clinical	Nail	25/Male	None	ND	Ahv
	IFRC 1173	MF185029	Clinical	Nail	46/Male	Diabetes	ND	Ahv
	IFRC 1174	MF185030	Clinical	Nail	46/male	ND	ND	Ahv
	IFRC 1175	MF185031	Clinical	Nail	27/Male	None	ND	Ahv
	IFRC 1177	MF185032	Clinical	Nail	27/Male	None	ND	Ahv
	IFRC 1178	MF185033	Clinical	Nail	53/Female	None	ND	Ahv
	IFRC 1179	MF185034	Clinical	Nail	53/Female	Diabetes	ND	Sar
	IFRC 1180	MF185035	Clinical	Nail	52/Male	Diabetes	ND	Sar
	IFRC 1181	MF185036	Environmental	Air				Teh
	IFRC 1182	MF185037	Environmental	Air				Teh
	IFRC 1282	MF185038	Clinical	Biopsy (Lung)	45/Male	Kidney transplant/ Diabetes	ITC/VRC	Sar
	IFRC 1283	MF185039	Environmental	Soil				Ahv
	IFRC 1284	MF185040	Environmental	Soil				Ahv
	IFRC 1286	MF185041	Environmental	Soil				Ahv
	IFRC 1287	MF185042	Environmental	Air				Ahv
	IFRC 1288	MF185043	Environmental	Soil				Teh
	IFRC 1290	MF185045	Clinical	BAL	45/Male	Asthma	ND	Teh
	IFRC 1291	MF185046	Environmental	Air				Ahv
	IFRC 1292	MF185047	Environmental	Air				Ahv
	IFRC 1296	MF185048	Clinical	Sputum	35/Female	COPD	ND	Teh
	IFRC 1516	MF185049	Clinical	Nail	63/Male	Diabetes	ND	Sar
	IFRC 1517	MF185050	Clinical	Nail	46/Female	None	ND	Sar
	IFRC 1519	MF185052	Environmental	Soil				Shi
	IFRC 1520	MF185053	Environmental	Soil				Shi
	IFRC 1521	MF185054	Environmental	Soil				Shi
	IFRC 1522	MF185055	Environmental	Soil				Shi
	IFRC 1524	MF185057	Clinical	Sputum	52/Male	Bronchitis	ND	Teh
	IFRC 1531	MF185059	Environmental	Soil				Ahv
	IFRC 1532	MF185060	Environmental	Soil				Ahv

48

49 50

52

TABLE 1 (Continued)

Species nr (%)	IFRC nr	Beta Tubulin accession nr	Source	Specimen	Age/Sex	Underlying condition	First-line therapy	Origin
	IFRC 1535	MF185062	Environmental	Soil				Ahvaz
	IFRC 1537	MF185063	Environmental	Soil				Ahvaz
	IFRC 1539	MF185065	Environmental	Soil				Ahvaz
	IFRC 1541	MF185066	Environmental	Soil			-	Ahvaz
	IFRC 1542	MF185067	Environmental	Soil			-	Ahvaz
	IFRC 1543	MF185068	Environmental	Soil				Ahvaz
	IFRC 1544	MF185069	Environmental	Soil		-		Ahvaz
	IFRC 1546	MF185070	Environmental	Soil		-	-	Ahvaz
	IFRC 1547	MF185071	Environmental	Soil		-	-	Ahvaz
	IFRC 1548	MF185072	Environmental	Soil		-	-	Ahvaz
	IFRC 1549	MF185073	Environmental	Soil		-		Ahvaz
	IFRC 1603	MF185074	Environmental	Soil				Ahvaz
	IFRC 1604	MF185075	Environmental	Soil				Ahvaz
	IFRC 1605	MF185076	Environmental	Soil		×		Ahvaz
	IFRC 1606	MF185077	Environmental	Soil	-	_		Ahvaz
	IFRC 1607	MF185078	Environmental	Soil				Ahvaz
	IFRC 1608	MF185079	Environmental	Soil	-			Ahvaz
	IFRC 1609	MF185080	Environmental	Soil	, V			Ahvaz
	IFRC 1613	MF185083	Environmental	Soil				Ahvaz
	IFRC 1614	MF185084	Environmental	Soil				Ahvaz
	IFRC 1665	MF185088	Clinical	Sputum	32/Male	Asthma	ND	Tehra
	IFRC 1685	MF185089	Clinical	BAL	51/Female	Lymphoma	ND	Mashl
	IFRC 1686	MF185090	Clinical	BAL	43/Female	ND	ND	Mashl
	IFRC 1687	MF185091	Environmental	Air				Mashl
A. citrinoterreus,	IFRC 493	MF185011	Environmental	Air				Sari
15(18.5%)	IFRC 1127	MF185012	Clinical	Biopsy	27/Male	Liver transplantation	AMB	Shiraz
	IFRC 1128	MF185013	Clinical	BAL	ND	COPD	AMB	Tehra
	IFRC 1129	MF185014	Clinical	Nail	21/Female	ND	ND	Sari
	IFRC 1130	MF185015	Clinical	Sinus	31/Female	None	ND	Tehra
	IFRC 1289	MF185044	Environmental	Soil				Tehra
	IFRC 1518	MF185051	Clinical	BAL	32/Female	Asthma	ND	Tehra
	IFRC 1523	MF185056	Clinical	Sputum	21/Female	Asthma	ND	Tehra
	IFRC 1525	MF185058	Clinical	Sputum	35/Female	Pneumonia	ND	Tehra
	IFRC 1538	MF185064	Environmental	Soil				Ahvaz
	IFRC 1610	MF185081	Environmental	Soil				Ahvaz
	IFRC 1612	MF185082	Environmental	Soil				Ahvaz
	IFRC 1662	MF185085	Clinical	Sputum	31/Female	Asthma	ND	Tehra
	IFRC 1663	MF185086	Clinical	Sputum	42/Male	Asthma	ND	Tehra
	IFRC 1664	MF185087	Clinical	Sputum	57/Male	Asthma	ND	Tehra

AMB, amphotericin B; ITC, itraconazole; VRC, voriconazole; BAL, Bronchoalveolar lavage; COPD, chronic obstructive pulmonary disease; IFRC, Invasive Fungi Research Center; ND, not determined.

 $_{51}$ $\,$ to triazoles. Basically, the GM MIC value of luliconazole against all

A. terreus isolates was >2 log2 dilutions lower than that of lanocona-zole.

53 1.2% (1/81) of the isolates showed amphotericin B MICs of \geq 4 µg/

mL. None of the isolates had MICs of >2 μ g/mL for voriconazole and itraconazole or >0.5 μ g/mL for posaconazole. All *A. terreus* iso-lates had low MECs of caspofungin (MEC range, 0.004--0.016 μ g/mL;

Strains and	MICs or MECs (µ	ւց/mL)													
antifungal drugs	Range	MIC50/MIC90	G mean	≤0.008	0.016	0.031	0.063	0.125	0.25	0.5	1	2	4	8 ≥16	≥64
Clinical (n = 36)															
AmB	0.252	1/2	0.8642						1	7	24	4			
FLU	64>64	64/64	64												36
ITC	0.0162	0.125/0.5	0.1321		3	5	3	10	9	3	2	1			
VRC	0.251	0.25/1	0.3762						18	13	5				
POS	0.1250.5	0.125/0.5	0.1815					25	7	4					
ISA	0.00160.125	0.031/0.063	0.0339		15	4	14	3							
LCZ	0.0010.031	0.016/0.016	0.0078	17	15	4									
LLCZ	0.0010.031	0.001/0.008	0.0019	35		1									
TER	0.0021	1/1	0.2646	5	1	2	2			9	19				
CFG	0.0040.016	0.008/0.008	0.008	35	1										
AFG	0.008	0.008/0.008	0.008	36											
MFG	0.0080.016	0.008/0.016	0.0093	27	9										
Environmental (n =	45)														
AmB	0.1254	2/2	1.234					1	2	6	13	22	1		
FLU	64>64	64/64	64												45
ITC	0.0160.5	0.125/0.25	0.1427		1	5	3	16	17	3					
VRC	0.1251	0.5/0.5	0.3407					5	16	23	1				
POS	0.0310.5	0.125/0.5	0.1753			4		25	5	11					
ISA	0.0010.063	0.031/0.063	0.0218	5	15	11	14								
LCZ	0.0010.031	0.001/0.031	0.0026	35	4	6									
LLCZ	0.0010.016	0.001/0.004	0.0014	44	1										
TER	0.0021	0.031/1	0.0423	13	5	9	1		1	9	7				
CFG	0.0040.008	0.008/0.008	0.0077	45											
AFG	0.008	0.008/0.008	0.008	45											
MFG	0.0080.016	0.008/0.016	0.0086	42	3										
All isolates (n = 81)															
AmB	0.1254	1/2	1.050					1	3	13	37	26	1		
FLU	64>64	64/64	64												81
ITC	0.0162	0.125/0.25	0.1379		4	10	6	26	27	5	2	1			
VRC	0.1251	0.5/0.5	0.3564					5	34	36	6				

TABLE 2 In vitro activity of amphotericin B and 11 comparators antifungal agents against clinical and environmental Aspergillus terreus species complex

VAEZI et al.

(Continues)

5

TABLE 2 (Continued)

Strains and	MICs or MECs (ıg/mL)														
antifungal drugs	Range	MIC50/MIC90	G mean	≤0.008	0.016	0.031	0.063	0.125	0.25	0.5	1	2	4	8	≥16	≥64
POS	0.0310.5	0.125/0.5	0.1781			4		50	12	15						
ISA	0.0010.125	0.031/0.063	0.0266	5	30	14	29	3								
LCZ	0.0010.031	0.008/0.031	0.0042	52	19	10										
LLCZ	0.0010.031	0.001/0.008	0.0016	79	1	1										
TER	0.0021	0.5/1	0.0972	18	6	11	3		1	18	24					
CFG	0.0040.016	0.008/0.008	0.0078	80	1											
AFG	0.008	0.008/0.008	0.008	81												
MFG	0.0080.016	0.008/0.016	0.0089	69	12											
A. terreus sensu str	<i>icto</i> (n = 66)															
AmB	0.1254	1/2	0.9792					1	3	13	30	18	1			
FLU	64	64/64	64													66
ITC	0.0161	0.125/0.5	0.1332		3	8	6	21	21	5	2					
VRC	0.1251	0.5/0.5	0.3462					5	28	30	3					
POS	0.0310.5	0.125/0.5	0.1576			4		40	8	14						
ISA	0.0040.125	0.031/0.063	0.0287	5	25	10	24	2								
LCZ	0.0010.031	0.008/0.031	0.0048	42	16	8										
LLCZ	0.0010.031	0.001/0.008	0.0018	64	1	1										
TER	0.0011	0.5/1	0.1192	12	5	10	1		1	15	22					
CFG	0.0040.008	0.008/0.008	0.0079	66												
AFG	0.008	0.008/0.008	0.008	66												
MFG	0.0080.016	0.008/0.016	0.0086	59	7											
A. citrinoterreus (n =	= 15)								\sim							
AmB	12	2/2	1.4472							\sim	7	8				
FLU	64	64/64	64													15
ITC	0.0162	0.125/0.5	0.1504		1	2		5	5	1		1				
VRC	0.251	0.5/1	0.4352						6	6	3					
POS	0.1250.5	0.125/0.25	0.1649					10	4	1						
ISA	0.0160.125	0.031/0.063	0.0329		5	5	4	1								
LCZ	0.0010.031	0.008/0.016	0.0045	10	3	2										
LLCZ	0.0010.031	0.001/0.008	0.0019	14		1										
TER	0.0011	0.063/1	0.0691	5	1	1	2			3	4					

(Continues)

2

4

5

6

7

9

10

11

12

13

15

16

17

18

19

21

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37 38 39

40

41

42 43

44

45

46

47

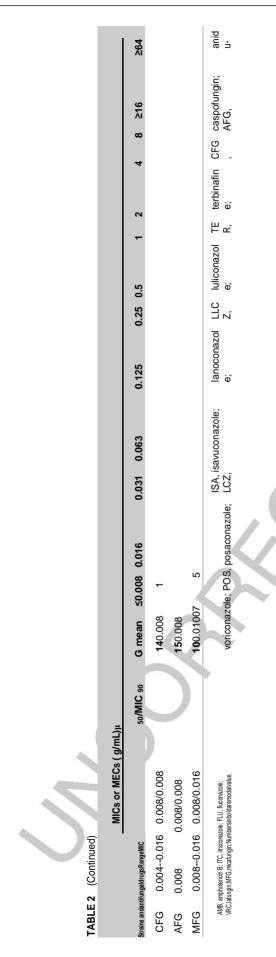
48

49

51

52

53



👷 mycoses — WILE

MEC90, 0.008 μ g/mL), micafungin (MEC range, 0.008--0.016 μ g/mL; MEC90,0.016 μ g/mL) and anidulafungin (MEC range, 0.008 μ g/mL; MEC90, 0.008 μ g/mL). There were significant differences in MICs of terbinafine (*P* < .0001) and amphotericin B (*P* < .002) between clinical and environmental isolates. However, no significant differences were found in the activities of terbinafine, amphotericin B, lanoconazole, luliconazole and micafungin between *A. terreus* and *A. citrinoterreus* (*P* > .05).

3.1 | Patient isolates

In all, 36 A. terreus isolates were collected in six tertiary care centres from five different cities. The mean age was 59.2 ± 14.2 years and 21 patients (58.3%) were male. In total, 11 (30.5%) of 36 patients were classified into the colonised group and 25 (69.6%) of 36 into the aspergillosis group. Among the patients with aspergil-losis, IA was diagnosed in 6 (16.7%), aspergilloma in 2 (5.6%), CPA in 3 (8.3%) and onychomycosis in 14 (38.9%) patients. According to diagnostic criteria, two cases out of six IA patients were classi-fied as proven IA and the other four cases identified as probable IA. The risk factors for IA patients were associated with COPD (n = 2), solid organ transplants (n = 2), lymphoma (n = 1) and his-tory of tuberculosis (n = 1). Due to the lack of available treatment data for all patients, we were unable to analyse outcome. Among three IA patients with treatment information, two received ampho-tericin B alone and one was treated with amphotericin B followed by voriconazole. Five of 6 patients with IA had a fatal outcome within 3--5 weeks of diagnosis. One of five patients died of other causes not related to IA. All the three cases with CPA received itra-conazole for 2--18 weeks and two patients died during treatment. Two patients with aspergilloma were treated with a combination of surgery and antifungal therapy (the first one received itraconazole alone and the second itraconazole for 1 week followed by voricon-azole for 3 months).

4 | DISCUSSION

Aspergillus citrinoterreus as a new species was found in patients with (n = 2) and without IA. The majority of *A. citrinoterreus* had been isolated from sputum and was regarded as coloniser (33.3%). It is difficult to make firm conclusions of this new species because of the lack of a comprehensive patient history and laboratory data.

In the present study, 98.8% (80/81) of all isolates showed amphotericin B MICs of<ECV (4 μ g/mL). Although several studies have shown that *A. terreus* species complex has intrinsic resistance to amphotericin B, some reports described *A. terreus* isolates with

4,17-20 low MICs. The reasons for the difference in amphotericin B

susceptibility patterns between studies remain unknown, but it has been speculated that genetic and species diversity in *Terrei* sec-tion may play an important role. However, Tortorano et al²¹ and

Neal et al²² were unable to show a relationship between low amphotericin B MICs and particular genotypes. Interestingly, in our

7

WILEY- mycose

8

study, a large number of A. terreus sensu stricto isolates were found 1 with low amphotericin B MICs. The data in the present study are in 2 concordance with a study by Risslegger et al.²³ who reported low 3 Δ amphotericin MICs (MIC range, 0.25--0.5 mg/L) in 6.3% A. terreus sensu stricto. Investigation of the impact of clinical, environmental 5 and even ecological factors on various amphotericin B susceptibil-ity 6 profiles in different countries can be considered. In the present report, 7 terbinafine has shown low in vitro activity against these isolates, 8 consistent with data published by Garcia--Effron et al²⁴ with MIC 9 range (0.003--4 µg/mL) and MIC90 (1 µg/mL). However, Fernandez 10 et al. reported lower values (MIC range, 0.003--0.25 µg/ mL; MIC90, 11 0.25 μ g/mL).¹⁷ The variable methodologies employed in previously 12 published studies, the failure of molecular identifica-tion and the 13

emergence of cryptic species may be the main rea-sons associated 14 with these differences. A prospective international 15 16 A. terreus analysis with 370 cases from 21 countries showed low

susceptibility to amphotericin B against the majority of cryptic spe-17 cies, the amphotericin B profiles of A. terreus on a worldwide scale is 18 still poorly understood and more research is needed.²³ Overall, in 19 20 vitro susceptibility testing can help to select an appropriate therapy 21 and to improve the management of patients infected with A. ter-reus. Susceptibility trends of A. terreus in surveillance studies can aid this 22 objective. 23

ACKNOWLEDGEMENTS

This research was financially supported by a grant of Mazandaran University, Faculty of Medicine, Sari, Iran (no. 2206) which we grate-28 fully acknowledge.

CONFLICT OF INTEREST

During the past 5 years, E.D has received research grants from MSD and Gilead; travel grants from Gilead, MSD, Pfizer and Astellas, and speaker's fee from Gilead, MSD and Astellas. The authors have no conflicts of interest regarding the content of this article.

ORCID

40	Tahereh Shokohi	http://orcid.org/0000-0003-3094-8436
41	Ferry Hagen	http://orcid.org/0000-0002-5622-1916
42 43	Hamid Badali	http://orcid.org/0000-0002-6010-8414

44 45

46

24 25

26 27

29 30 31

32 33

34

35

36 37 38

39

- REFERENCES
- 1. Hagiwara D, Watanabe A, Kamei K, Goldman GH. Epidemiological $^{4/}$ and genomic landscape of azole resistance mechanisms in Aspergillus 48

Fungi. Front Microbiol. 2016;7:1382.

2. Scorzoni L, de Paula E, Silva AC, Marcos CM, et al. Antifungal therapy: 49

50 new advances in the understanding and treatment of mycosis. Front Microbiol. 2017;8:36. https://doi.org/10.3389/fmicb.2017.00036

3. Pastor FJ, Guarro J. Treatment of Aspergillus terreus infections: a 52

- clinical problem not yet resolved. Int J Antimicrob Agents. 2014;44: 53 281-289.
- Blum G, Hörtnagl C, Jukic E, et al. New insight into amphoteri-cin B resistance in Aspergillus terreus. Antimicrob Agents Chamother 2013:57:1583-1588
- 5. Deak E, Wilson SD, White E, Carr JH, Balajee SA. Aspergillus terreus accessory conidia are unique in surface architecture, cell wall compo-sition and germination kinetics. PLoS One. 2009;4:e7673. https://doi. org/10.1371/journal.pone.0007673
- 6. Lass-Flörl C. Susceptibility testing in Aspergillus species complex. Clin Microbiol Infect. 2014;20(Suppl 6):49-53.
- 7. Al-Quadeib BT, Radwan MA, Siller L, et al. Therapeutic monitoring of amphotericin B in Saudi ICU patients using UPLC MS/MS assay.

Biomed Chromatogr. 2014;28:1652-1659.

- Arendrup MC, Jensen RH, Grif K, et al. *In vivo* emergence of *Aspergillus terreus* with reduced azole susceptibility and a *Cyp51*a M217I alter-ation. *J Infect Dis.* 2012;206:981-985.
- Chowdhary A, Kathuria S, Xu J, et al. Clonal expansion and emergence of environmental multiple--triazole--resistant *Aspergillus fumigatus* strains carrying the TR(3)(4)/L98H mutations in the *cyp51A* gene in India. *PLoS One*. 2012;7:e52871.
- Hong SB, Go SJ, Shin HD, Frisvad JC, Samson RA. Polyphasic tax-onomy of *Aspergillus fumigatus* and related species. *Mycologia*. 2005;97:1316-1329.
- Clinical and Laboratory Standards Institute. Reference Method For Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi, 2nd ed; approved standard. CLSI document M38–A2. Wayne, PA, USA: Clinical and Laboratory Standards Institute; 2008.
- Espinel-Ingroff A, Fothergill A, Fuller J, Johnson E, Pelaez T, Turnidge J. Wild--type MIC distributions and epidemiological cutoff values for caspofungin and *Aspergillus* spp. for the CLSI broth microdilu-tion method (M38--A2 document). *Antimicrob Agents Chemother*. 2011;55:2855-2859.
- Espinel-Ingroff A, Cuenca-Estrella M, Fothergill A, et al. Wild--type MIC distributions and epidemiological cutoff values for amphotericin B and *Aspergillus* spp. for the CLSI broth microdilution method (M38--A2 document). *Antimicrob Agents Chemother*. 2011;55: 5150-5154.
- Ascioglu S, Rex JH, DePauw BE, et al. Defining opportunistic invasive fungal infections in immunocompromised patients with cancer and hematopoietic stem cell transplants: an international consensus. *Clin Infect Dis.* 2002;34:7-14.
- Kathuria S, Sharma C, Singh PK, et al. Molecular epidemiology and in--vitro antifungal susceptibility of *Aspergillus terreus* species complex isolates in Delhi, India: evidence of geneticdiversity by amplified fragment length polymorphism and microsatellite typing. *PLoS One*. 2015;10:e0118997.
- Hubka V, Kubatova A, Mallatova N, et al. Rare and new etiological agents revealed among 178 clinical *Aspergillus* strains obtained from Czech patients and characterized by molecular sequencing. *Med Mycol.* 2012;50:601-610.
- Fernández MS, Rojas FD, Cattana ME, et al. *In vitro* activities of am-photericin B, terbinafine, and azole drugs against clinical and environ-mental isolates of *Aspergillus terreus sensu stricto*. *Antimicrob Agents Chemother*. 2015;59:3619-3622.
- Meletiadis J, Antachopoulos C, Stergiopoulou T, Pournaras S, Roilides E, Walsh TJ. Differential fungicidal activities of amphotericin B and voriconazole against *Aspergillus* species determined by microbroth methodology. *Antimicrob Agents Chemother*. 2007;51: 3329-3337.
- Gomez-Lopez A, Garcia-Effron G, Mellado E, Monzon A, Rodriguez-Tudela JL, Cuenca-Estrella M. *In vitro* activities of three licensed antifungal agents against spanish clinical iso-lates of *Aspergillus* spp. *Antimicrob Agents Chemother*. 2003;47: 3085-3088.
- 20. Misra R, Malik A, Singhal S. Comparison of the activities of amphotericin B, itraconazole, and voriconazole against clinical and

1	environmental isolates of Aspergillus species. Indian J Pathol Microbiol.	24. Garcia-Effron G, Gomez-Lopez A, Mellado E, Monzon A, Rodriguez-
2	2011;54:112-116.	Tudela JL, Cuenca-Estrella M. In vitro activity of terbinafine against
3	 Tortorano AM, Prigitano A, Dho G, et al. In vitro activity of amphoter-icin B against Aspergillus terreus isolates from different countries and 	medically important nondermatophyte species of filamentous fungi.
4 5	against Aspergillus terreus isolates from different countries and regions. J Chemother. 2008;20:756-757. 22. Neal CO, Richardson AO, Hurst SF, et al. Global population structure	J Antimicrob Chemother. 2004;53:1086-1089.
		How to cite this article: Vaezi A, Fakhim H, Arastehfar A,
6 7	of Aspergillus terreus inferred by ISSR typing reveals geographical sub- clustering. <i>BMC Microbiol.</i> 2011;11:203.	et al. In vitro antifungal activity of amphotericin B and 11
	23. Risslegger B, Zoran T, Lackner M, et al. A prospective interna-	comparators against Aspergillus terreus species complex.
8	tional Aspergillus terreus survey: an EFISG, ISHAM and ECMM joint	
9	study. Clin Microbiol Infect. 2017;23:776.e1-776.e5. https://doi.	Mycoses. 2017;00:1–9. <u>https://doi.org/10.1111/myc.12716</u>
10	org/10.1016/j.cmi.2017.04.012	
11		
12		
13		
14		
15		
16		
17		
18		X
19		
20		
21		
22		
23		
24		
25	A	
26		
27 28		*
29		
30		
31		
32		
33		
34		
35		
36		
37		
38		
39		
40		
41		
42		
43		
44		
45		
46		
47		
48		
49		
50		
51		
52		
53		

-WILE